

IBC Meeting notes 9/2/26

Present: Crista Wadsworth, Paul Craig, MaryAnne Courtney, Sarah Klein, Emily Coon, Cindy White, Phil VanChieri, Dawn Carter, Stefan Schulze, Kim Corbett, Jennifer Harman, Zhongwang Dou

- 1) **Announcement:** Crista is stepping down as the Chair of the IBC but will remain an active member of the committee. After several discussions, Stefan Schulze was asked and has agreed to take over as the Chair of the IBC. Stefan's background is similar to Crista's and they currently share lab space in Brown Hall. Stefan is open to feedback and welcomes suggestions for improvement opportunities. The committee thanks Crista for her guidance and contributions and welcomes Stefan! We look forward to continuing to work with both of you.

- 2) **Protocol review:** Zhongwang Dou – Animal blood clot; Bovine, porcine or ovine heart organ tissue
 - a. Zhongwang is an assistant professor in BME studying fluid mechanics relative to biological components. He studies fluid flow in the human body using a mechanical representation of the circulatory system and cameras to view blood clot movement in the system.

 - b. **Registration:**
 - i. No changes were required.

 - c. **SOP:**
 - i. The following details were discussed: (highlighted items need to be added to the SOP)
 1. The system will be a closed loop that is in the open lab. It will not fit into a hood. Eye and face protection, along with a lab coat and gloves will be required and **need to be called out in the SOP**. While there is little chance of a spill or splash as clotted blood will be used, precautions must be taken.
 2. The pressure in the system will be 120 mmHg up to 240 to mimic human blood pressure.
 3. External cleaning of the system will be done with 70% ethanol. Internal cleaning will be done with 10% bleach. All cleaning and rinse fluids will be **collected and discarded in the appropriate waste stream**.
 4. Heart organ: will be placed in the flow loop to see how the clot flows through it.
 - a. The **valves will be prepped in a biosafety cabinet** and treated as though they are infected or contaminated.

b. All blood and tissues transported will be placed in appropriate packaging for intrafacility transport based on DOT guidelines: It

is also common to need to move samples or cultures between laboratories, between floors in a building, or by walking samples between buildings. When a sample needs to be moved, care should be taken to minimize the transport through public and office areas. Avoid passenger elevators when possible, using stairs and freight elevators instead. It is recommended that the sample(s) be placed in a sealable bag or container to provide primary leak-proof containment. Place absorbent in the bag or container to absorb any spilled material in the event of a loss. Place the sealed bag or container in a durable, rigid outer container for transport. Disinfect the exterior of the outer container as appropriate depending on the risk posed by the material to be transported. PPE to be worn during transit is based on the institution's risk assessment.

3) Bolaji Thomas: Dried blood spots – malaria

a. Registration comments

- i. Section B1 indicates that the samples are human blood infected with plasmodium falciparum but section K says that they are uninfected. Are the samples from individuals who have malaria or no?
- ii. Section B1a – specify who the collaborator is
- iii. Dried plasmodium falciparum cannot survive, but have the samples been tested for other pathogens? How do we know that nothing else is present? (Hep B, C and HIV were specifically mentioned)
- iv. Section K states 'if positive by PCR'. What are they being screened for?
- v. All samples need to be treated as infectious.

b. SOP Comments – DNA Extraction

- i. The SOP doesn't describe waste handling or PPE. Add details for both.
- ii. Will ethidium bromide be used? If so, handling and disposal must be included.
- iii. How is the DNA concentration being measured?
- iv. Will undergrads be working alone? The committee feels that this is acceptable if a buddy is present and both have had the appropriate training.

c. MTA – there does not appear to have been a Materials Transfer Agreement (MTA) in place to transport these samples to RIT. When biological samples are shipped from a collaborator to RIT, an MTA should be in place to let RIT know that they are coming and to sign off on them.

d. Packaging for shipping – The packaging that these samples were in, with no secondary containment and an envelope that could be damaged during shipping was not adequate. There are concerns that if the packaging was damaged and the papers got wet, that the samples could potentially be hazardous. The DOT has specific requirements for how biological samples need to be packed and labeled for air transport. The full guidelines will be provided to the committee as well as to the PI.

4) Discussion –

- a. MTA – Is one required to transfer samples? Should the OTA be contacted for guidance? How do we enforce this?
- b. Suggested edits to the registration:

- i. Add a text box for funding sources vs Is it NIH funded?
 - ii. Add the question: Do you have an MTA for your material?
 - iii. Can the department chair be brought into the conversation to enforce the MTA?
 - iv. Add a line for a grant or SRS# to be added.
- c. Todd Camenisch has asked for an update on the status of projects in CHST. Cindy will be putting together an email in the next week to let him know what's currently happening.

I think that's everything. Please let me know if I missed anything or if changes are needed.