

9/25/2023 IBC Meeting

Present:

Jennifer Liedkie, Karin Wuertz-Kozak, Phil Van Chieri, Cindy White, Dawn Carter, Gary Skuse, Sarah Klein, Kim Corbett, Paul Craig, Jennifer Harman

Experimental design needs to be in the project registration form

-Clarify what is happening at RIT vs Africa; The work in Africa when it comes to tissue sampling, preserving, and preparation for shipping needs to be clearer (who's shipping it, etc.) and brief. The RIT work needs to be much more defined.

-RNA preservation of tissue samples through to Bullet 3, then Bullet 5 says tissue samples will be shipped

-Shipped to PI – each PI needs to be named explicitly

-Got a few details on RNALater, RNase inhibitor, unsure about proteins. It's used for RNA and protein insolation. What's in it? SDS sheet - Ammonium sulfate. How does it RNase deactivate protein/RNA? The committee needs to learn about it.

-HOW WILL THE SAMPLES BE ON CAMPUS preserved vs. fixed, form and function.

Sponsored Research- will get the USDA permit. IBC ask for permit application. Prions are still a concern for USDA. Still working on getting it- status?

Safety protocol

-Still things about "class"

-Need tissue specific safety, and tissue waste disposal (biohazard, but what form) clarification

-Fixed- means specific no longer hazardous or dangerous, but that changes biolevel. Formalin fixing for example. Clarify if that's RNALater or truly fixed. Preserved is not fixed. Clarify in protocol.

IBC letters will be incoming, hopefully October.